(wileyonlinelibrary.com) DOI 10.1002/bio.2486

Microwave-assisted one-step green synthesis of amino-functionalized fluorescent carbon nitride dots from chitosan

Deli Xiao,^a Danhua Yuan,^a Hua He^{a,b*} and Jinrong Lu^c

ABSTRACT: We present an economical, facile and effective microwave pyrolysis approach to synthesize highly aminofunctionalized fluorescent carbon nitride dots (CNDs). The formation and the functionalization of CNDs was accomplished simultaneously through the dehydration of chitosan. It is suggested that these CNDs have good water solublility and exhibit strong fluorescence. Copyright © 2013 John Wiley & Sons, Ltd.

 \Box Supporting information may be found in the online version of this article.

Keywords: microwave; green synthesis; fluorescent carbon nitride dots; chitosan

Introduction

In recent years, fluorescent semiconductor quantum dots (QDs) have been a common subject of academic research because of their unique optical and biochemical features (1–6). However, the use of heavy metals, which are essential elements in these conventional semiconductors, is restricted because of concerns about their toxicity, stability and environmental impact (8,7). Therefore, the fabricatation of benign nanomaterials with similar optical properties is an interesting challenge inspiring intensive research. In recent years, a new type of visible emitter has been reported, based exclusively on carbon dots (CDs) (9–12) or carbon nitride dots (CNDs) (13,14). CDs and CNDs seem to be a promising alternative to traditional toxic metal-based semiconductor QDs in many fields of applications due to their stable photoluminescence (PL), strong fluorescence, broad excitation spectra, tunable emission spectra, low cytotoxicity and excellent biocompatibility.

Approaches to synthesizing eco-friendly fluorescent CDs or CNDs can be generally classified into two main groups: top-down and bottom-up (15). Top-down methods are used to etch a larger carbon structure into individual nanoparticles, such as arcdischarge single-walled carbon nanotubes (16), laser ablation of graphite (9), electrochemical oxidation of graphite and multiwalled carbon nanotubes (17,18), carbonizing polymerized resols on silica spheres (19), chemical oxidation soots of candles, natural gas, commercially activated carbon and lampblack (20–22) and chemical oxidation of oxide graphene (23). By contrast, bottomup methods are used to form nanoparticles from molecular precursors, such as chemical and thermal oxidation or in the microwave pyrolysis of carbonaceous compounds (14,24–28). However, most of these synthesis methods involve expensive starting materials, devices that consume a great deal of energy, intricate processes, and the as-synthesized CDs or CNDs are typically oxidized by strong acid and further surface-passivated by a diamine-terminated organic molecule to improve the water solubility of the CNDs and modify the PL properties. Therefore, building an economical, facile, effective and green synthetic route to produce strong fluorescent CDs or CNDs on a large scale for practical application remains critical.

Herein, we present an economical, facile and effective microwave pyrolysis approach for the synthesis of highly aminofunctionalized fluorescent CNDs. A characteristic feature of this one-step approach is that the formation and the functionalization of CNDs was accomplished simultaneously through the dehydration of chitosan; neither a strong acid solvent nor surface modification reagent is needed. The synthetic process occurs in a domestic microwave oven using inexpensive chitosan as the carbon source and has the advantage of being inexpensive and completely 'green'.

Experimental

Preparation of fluorescent CNDs

In a typical procedure, amino-functionalized fluorescent CNDs were synthesized as follows: 0.5 g chitosan was dissolved in 10 mL water to form a homogeneous solution, which was then put into a domestic microwave oven (700 W) and heated for 9.5 min. The color-changed solution was centrifuged at 13,000 rpm for 30 min to remove the less-fluorescent deposit, and dialyzed against pure water through a dialysis membrane for 4 days to remove residual chitosan. Finally, amino-functionalized fluorescent CNDs were dried in a vacuum at 65 \degree C overnight.

* Correspondence to: Hua He, Deparment of Analytical Chemistry, China Pharmaceutical University, Nanjing 210009, China. E-mail: dochehua@163.com

- b Key Laboratory of Drug Quality Control and Pharmacovigilance, Ministry of Education, China Pharmaceutical University, Nanjing 210009, China
- Department of Organic Chemistry, China Pharmaceutical University, Nanjing 210009, China

a Department of Analytical Chemistry, China Pharmaceutical University, Nanjing 210009, China

Characterization methods

UV–vis absorption was characterized using a UV1800 UV–vis spectrophotometer (Shimadzu Corporation, Japan). PL emission measurements were performed using a RF-5301PC fluorescence spectrophotometer (Shimadzu Corporation). The morphology of the as-synthesized nanoparticles was studied using a FEI Tecnai G2 F20 transmission electron microscope (TEM). Elemental analysis was carried out using an Elementar Vario ELIII. The surface groups on CNDs were measured with a 8400 s FT-IR spectrometer (Shimadzu Corporation). X-Ray photoelectron spectroscopy (XPS) analysis was measured on an ESCALAB MK-II X-ray photoelectron spectrometer.

Quantum yield measurements

The quantum yield (Φ) of CNDs was measured by comparing the integrated PL intensities and the absorbency values against the reference quinine sulfate (QS). QS (literature $\Phi = 0.54$) was dissolved in 0.1 M H₂SO₄ (refractive index, η , of 1.33) and the CNDs were dissolved in distilled water (η = 1.33).

$$
\varPhi=\varPhi_R\times\frac{I}{I_R}\times\frac{A_R}{A}\times\frac{\eta^2}{\eta_R{}^2}
$$

where Φ is the quantum yield, *I* is the measured integrated emission intensity, η is the refractive index and A is the optical density. The subscript R refers to the reference fluorophore of known quantum yield.

Results and discussion

As shown in Scheme 1, in a typical procedure, amino-functionalized fluorescent CNDs were synthesized as follows: an appropriate amount of chitosan was dissolved in water to form a homogeneous solution, which was then put into a domestic microwave oven (700 W) and heated for different lengths of time. The color-changed solution was centrifuged and dialyzed to remove less-fluorescent deposit and residual chitosan. Finally, a clear yellow–brown aqueous solution containing CNDs was obtained. The yield of fluorescent CNDs is \sim 6.4% and the sample shows good water solubility. Experiments revealed that the solubility of the CNDs

reached 60 mg/mL in water and there was no precipitation for months. Our preparation method presents a facile approach to producing highly amino-functionalized fluorescent CNDs on a large scale. The functional groups on its surface improve its water solubility and reduce its potential biotoxicity, which is essential for biologically motivated work.

Figure 1 depicts the UV–vis absorption and PL spectra for CNDs. The first absorption band was observed at 282 nm. When it was excited at 338 nm, the PL spectra showed a peak position at 440 nm, a 158 nm red shift from the first absorption peak. In addition, the fluorescent intensity gradually decreased with increasing excitation wavelength. Such an observation is similar to that of carbon dots, which may be attributed to the optical selectivity of differently sized nanoparticles (quantum effect) or different emissive traps on the CND surface or another mechanism altogether. In addition, abundant functional groups, such as carboxyl acids and amines, which are introduced simultaneously during the microwave pyrolysis, may introduce different defects onto the surface of CNDs, acting as excitation energy traps and leading to different PL properties.

The fluorescence quantum yield of CNDs is \sim 6.4% when the microwave pyrolysis time is 9.5 min with QS as a standard reference (Table S1). These values are comparable with previous

Figure 1. UV-vis absorption spectra and PL spectra for CNDs.

Scheme 1. A schematic illustration of the prepration procedure of CNDs by microwave pyrolysis of chitosan.

reports (13,29). Table S2 shows the PL quantum yield of CNDs for various microwave pyrolysis times. It can be seen that the PL quantum yield increased sharply when microwave pyrolysis time varied from 3.5 to 9.5 min, which indicated that the formation of CNDs is very fast and facile.

The TEM image (Fig. 2) showed that the nanoparticles were monodispersed and uniform with a spherical shape. The asprepared CNDs were well dispersed in narrow distributions of 4.6 ± 1.9 nm diameter with a microwave pyrolysis time of 9.5 min. As the time increased to 11.5 min, the diameter of the CNDs increased slightly and an aggregate phenomenon was observed. Thus, a microwave pyrolysis time of 9.5 min was chosen in the subsequent study. The chemical composition of these nanoparticles was further determined by collecting the corresponding energy-dispersed spectroscopy (EDS) results, as shown in Fig. S1. Peaks of C, N and O are observed, indicating that these nanoparticles are formed by chitosan and H_2O . Furthermore, peaks of F, S, Si and Cu are also observed, which originate from the glass substrate used for EDS analysis. Elemental analysis indicated that the composition of the CNDs was: C, 34.93 wt%; N, 6.43 wt%; H, 6.97 wt% and O, 51.67 wt%. It was found that after microwave carbonization the carbon and nitrogen content of the CNDs increased, which was mainly due to the loss of oxygen and hydrogen during dehydration of the chitosan (Table S3). The surface composition and elemental analysis for the overall composition of the resultant nanoparticles were characterized using XPS techniques. The XPS spectrum of the nanoparticles shown in Figs S2–S5 exhibited three peaks at 285, 401 and 532 eV, which were attributed to C1s, N1s and O1s, respectively. All these results indicated that the nanoparticles thus obtained were mainly composed of C, N, O and H.

As shown in Fig. 3, Fourier transform infrared (FT-IR) measurements of pure chitosan powder showed that the peaks at 3447 cm^{-1} were attributed to O-H and N-H stretching vibrations of amine groups. The peaks at 1644 cm^{-1} and 1593 cm^{-1} corresponded to N–H bending vibrations. For the CNDs, the result showed an increase in absorption of the amino group at 1634 cm^{-1} compared with chitosan. C-H vibrations at 1130–1064 cm^{-1} associated with the pyranose were almost lost. These distinctions may be caused by degradation of the chitosan chain and decomposition of the pyranose ring through dehydration. Similar results are also reported in the literature

Figure 3. FT-IR spectrum of (a) pure chitosan, (b) fluorescent CNDs.

(27). The zeta-potential measurements showed that the surface of the CNDs has the amine groups because it is positive when the pH was $<$ 6.6, suggesting that microwave pyrolysis of chitosan was an effective way to obtain amino-functionalized fluorescent CNDs.

Moreover, the impact of pH and ionic strength on the fluorescence intensity of CNDs was investigated (Fig. S6). The results indicated that the fluorescence intensity of CNDs changed slightly when the solution pH varied from 5.3 to 7.7, and even in an aqueous solution with a high ionic strength the fluorescence intensity of the CNDs was stable. These phenomena show the potential of the fluorescent CNDs to be good candidates for biological applications.

Conclusions

In conclusion, fluorescent CNDs were prepared by microwave pyrolysis derived from chitosan. The synthesized CNDs showed some benign properties such as higher PL efficiencies (6.4%), monodispersity and small diameter $(4.6 \pm 1.9 \text{ nm})$. In comparison with previous methods, this one-step 'green' process has no need for strong acid treatment or surface modification. In particular, the as-prepared CNDs exhibit excellent stability in biological media, and their luminescence intensity is also stable within the

Figure 2. TEM images and their size distributions for CNDs with different microwave pyrolysis time of (a) 9.5 min and (b) 11.5 min; scale bars: 50 nm.

physiological and pathological ranges for ionic strength or pH. The strong fluorescence and excellent water dispersion can be attributed to the abundant surface traps and functional groups. Combing their low cytotoxicity, low cost, green method of synthesis, ease of labeling and favorable optical properties, CNDs provide promising applications in biological labeling and biosensors.

SUPPORTING INFORMATION

Supporting information may be found in the online version of this article.

References

- 1. Selvan ST, Patra PK, Ang CY, Ying JY. Synthesis of silica-coated semiconductor and magnetic quantum dots and their use in the imaging of live cells. Angewandte Chemie-International Edition 2007;46: 2448–52.
- 2. Farias PMA, Santos BS, de Thomaz AA, Ferreira R, Menezes FD, Cesar CL, et al. Fluorescent II–VI Semiconductor Quantum Dots in Living Cells: Nonlinear Microspectroscopy in an Optical Tweezers System. The Journal of Physical Chemistry B 2008;112:2734–7.
- 3. Boeneman K, Delehanty JB, Susumu K, Stewart MH, Medintz IL. Intracellular Bioconjugation of Targeted Proteins with Semiconductor Quantum Dots. Journal of the American Chemical Society 2010;132:5975–7.
- 4. Petta JR, Johnson AC, Taylor JM, Laird EA, Yacoby A, Lukin MD, et al. Coherent manipulation of coupled electron spins in semiconductor quantum dots. Science 309:2005;2180–4.
- 5. Reithmaier JP, Sek G, Loffler A, Hofmann C, Kuhn S, Reitzenstein S, et al. Strong coupling in a single quantum dot-semiconductor microcavity system. Nature 2004;432:197–200.
- 6. Kroutvar M, Ducommun Y, Heiss D, Bichler M, Schuh D, Abstreiter G, et al., Optically programmable electron spin memory using semiconductor quantum dots. Nature 2004;432:81–4.
- 7. Lewinski N, Colvin V, Drezek R. Cytotoxicity of Nanoparticles. Small 2008;4:26–49.
- 8. Derfus AM, Chan WCW, Bhatia SN. Probing the cytotoxicity of semiconductor quantum dots. Nano Letters 2004;4:11–18.
- 9. Sun Y-P, Zhou B, Lin Y, Wang W, Fernando KAS, Pathak P, et al. Quantum-Sized Carbon Dots for Bright and Colorful Photoluminescence. Journal of the American Chemical Society 2006;128:7756–7.
- 10. Li H, He X, Kang Z, Huang H, Liu Y, Liu J, et al. Water-Soluble Fluorescent Carbon Quantum Dots and Photocatalyst Design. Angewandte Chemie International Edition 2010;49:4430–4.
- 11. Zong J, Zhu Y, Yang X, Shen J, Li C. Synthesis of photoluminescent carbogenic dots using mesoporous silica spheres as nanoreactors, Chemical Communications. Chemical Communications 2011;47:764–6.
- 12. Liu H, Ye T, Mao C. Fluorescent Carbon Nanoparticles Derived from Candle Soot. Angewandte Chemie International Edition 2007;46:6473–5.
- 13. Liu S, Wang L, Tian J, Zhai J, Luo Y, Lu W, et al. Acid-driven, microwave-assisted production of photoluminescent carbon nitride dots from N,N-dimethylformamide. RSC Advances 2011;1:951.
- 14. Liu S, Tian J, Wang L, Luo Y, Zhai J, Sun X. Preparation of photoluminescent carbon nitride dots from CCl4 and 1,2-ethylenediamine: a heat-treatment-based strategy. Journal of Materials Chemistry 2011;21:11726.
- 15. Baker SN, Baker GA. Luminescent Carbon Nanodots: Emergent Nanolights. Angewandte Chemie International Edition 2010;49:6726–44.
- 16. Xu X, Ray R, Gu Y, Ploehn HJ, Gearheart L, Raker K, et al. Electrophoretic Analysis and Purification of Fluorescent Single-Walled Carbon Nanotube Fragments. Journal of the American Chemical Society 2004;126:12736–7.
- 17. Zhao Q-L, Zhang Z-L, Huang B-H, Peng J, Zhang M, Pang D-W. Facile preparation of low cytotoxicity fluorescent carbon nanocrystals by electrooxidation of graphite. Chemical Communications 2008;5116.
- 18. Zhou J, Booker C, Li R, Zhou X, Sham T-K, Sun X, et al. An Electrochemical Avenue to Blue Luminescent Nanocrystals from Multiwalled Carbon Nanotubes (MWCNTs). Journal of the American Chemical Society 2007;129:744–5.
- 19. Bourlinos AB, Stassinopoulos A, Anglos D, Zboril R, Georgakilas V, Giannelis EP. Photoluminescent Carbogenic Dots. Chemistry of Materials 2008;20:4539–41.
- 20. Mochalin VN, Gogotsi Y. Wet Chemistry Route to Hydrophobic Blue Fluorescent Nanodiamond. Journal of the American Chemical Society 2009;131:4594–5.
- 21. Tian L, Ghosh D, Chen W, Pradhan S, Chang X, Chen S. Nanosized Carbon Particles From Natural Gas Soot. Chemistry of Materials 2009;21:2803–9.
- 22. Qiao Z-A., Wang Y, Gao Y, Li H, Dai T, Liu Y, et al. Commercially activated carbon as the source for producing multicolor photoluminescent carbon dots by chemical oxidation. Chemical Communications 2010;46:8812.
- 23. Sun X, Liu Z, Welsher K, Robinson JT, Goodwin A, Zaric S, et al. Nanographene oxide for cellular imaging and drug delivery. Nano Research 2008;1:203–12.
- 24. Zhao L, Baccile N, Gross S, Zhang Y, Wei W, Sun Y, et al. Sustainable nitrogen-doped carbonaceous materials from biomass derivatives. Carbon 2010;48:3778–87.
- 25. Liu C, Zhang P, Tian F, Li W, Li F, Liu W. One-step synthesis of surface passivated carbon nanodots by microwave assisted pyrolysis for enhanced multicolor photoluminescence and bioimaging. Journal of Materials Chemistry 2011;21:13163.
- 26. Zhu H, Wang X, Li Y, Wang Z, Yang F, Yang X. Microwave synthesis of fluorescent carbon nanoparticles with electrochemiluminescence properties. Chemical Communications 2009;5118.
- 27. Yang YH, Cui JH, Zheng MT, Hu CF, Tan SZ, Xiao Y, et al. One-step synthesis of amino-functionalized fluorescent carbon nanoparticles by hydrothermal carbonization of chitosan. Chemical Communications 2012;48:380–2.
- 28. Tian L, Song Y, Chang X, Chen S. Hydrothermally enhanced photoluminescence of carbon nanoparticles. Scripta Materialia 2010;62:883–6.
- 29. Liu S, Tian J, Wang L, Luo Y, Sun X. A general strategy for the production of photoluminescent carbon nitride dots from organic amines and their application as novel peroxidase-like catalysts for colorimetric detection of H2O2 and glucose. RSC Advances 2012;2:411.